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Back to the past—forever young: cutting-edge biochemical and microbiological tools for cultural heritage conservation

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1	Back to the past. Forever young: cutting-edge biochemical and
2	microbiological tools for cultural heritage conservation
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19 ABSTRACT

Ancient documents and milestones of human history such as manuscripts and textiles are 20 fragile and during aging undergo chemical, physical and biological deterioration. Among the different 21 causes of damage, also human intervention plays a role since some restoration strategies proved to be 22 23 transient and/or they generated further damage. Outdoor monuments undergo deterioration since they are exposed to pollution, weathering, microbial attack (giving rise to undesired pigmentation, 24 25 discoloration or true dissolution, corrosion, and overall decay) as well as men-made damage (i.e. graffiti). This review article reports the best fitting strategies used to restore wall paintings, outdoor 26 27 monuments, textiles and paper documents to their ancient beauty by employing "soft" bio-based approaches such as viable bacteria or suitable enzymes. 28

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Key-words: immobilized enzymes, biocleaning, caseinase, collagenase, viable bacteria, graffiti,
bioconsolidation

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34 INTRODUCTION

Artworks may undergo a number of degradation and deterioration events, which widely vary 35 depending on the specific artifact and the environment and conditions of conservation. These 36 parameters may be extremely different if we consider, for instance, a book conserved at controlled 37 temperature and humidity in a library or in a museum, or a stone statue or a cathedral, which are 38 constantly exposed to weathering, pollution, microbial colonization (extensively reviewed in Mazzoli 39 et al. 2018), vandalism acts, etc... It is worth reminding that damaging of artworks is sometimes the 40 effect of previous restoration interventions which underwent deterioration themselves during time, as 41 in the case of glues applied to consolidate wall paintings or ancient textiles (Beutel et al. 2002; Ferrari 42 et al. 2017). 43

Cleaning and/or restoration of artworks by biotechnological approach has been performed by 44 using enzymes or microorganisms or a combination of both strategies, depending on the specific 45 artifact and issue (Figs. 1, 2; Table 1). Bio-based methods have a number of advantages over more 46 consolidated techniques for artwork restoration (such as those using non-acqueous solvents, 47 bleaching and mechanical treatments), because of their lower impact on the environment, reduced 48 toxicity for operators and higher selectivity and safety for artworks themselves (Barbabietola et al. 49 2016). Enzymes are generally characterized by extremely high substrate specificity which allows high 50 selective choice depending on the "damaging material" (e.g. proteins, polysaccharides, lipids) to 51 treat/remove. Moreover, enzymes can be chosen whose catalytic activity is optimal at the most 52 53 suitable pH/T ranges for treating a certain artwork thus reducing the application time (Germinario et al. 2017). Enzymes have been used in aqueous formulations, with or without a gel as sorbent, and in 54 ionic liquids (Hrdlickova Kuckova et al. 2014). On the other hand, the use of enzymes may be limiting 55 56 because of the relatively high amounts required, their relatively high cost, the need for controlled application conditions (e.g. pH and T) and for skilled operators (Barbabietola et al. 2016). Therefore, 57 58 the use of microorganisms has sometimes been preferred, for instance when very resistant or complex deposit materials (i.e. mixture of heterogeneous substances) or very extended surfaces (e.g. the 59 surface of a cathedral) needed to be removed/treated. In addition, the use of living microorganisms is 60 necessary in the case of complex phenomena such as calcium carbonate deposition for the 61 62 bioconsolidation of stone material (Dhami et al. 2014). As compared to enzymatic strategies, the use of living microorganisms for biocleaning of artworks is certainly less expensive and may require less 63 64 controlled environmental conditions, although it is generally less selective (Webster and May 2006).

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67 One of the most common causes of biodeterioration of ancient papers (e.g. books, documents) and textiles that are preserved in museums, libraries and archives are glues employed, with specific 68 variations and modifications, to manufacturing or consolidating/restoring these artifacts 69 (Barbabietola et al. 2016; Ferrari et al. 2017). In the case of paper artworks, glues have been used in 70 manufacturing, such as for bonding and lining of prints, drawings, documents, which were mounted 71 (partly or completely) on secondary support by means of glue spots, as well as for restoration 72 73 (Barbabietola et al. 2016). As regards historical or ethnographic textiles, glues have mainly been used 74 for restoration purposes, e.g. to fasten them to textiles or to solid (paper or wood) supports (Ahmed and Kolisis 2011). In the past, glues of either vegetal (i.e. starch) or animal (i.e. collagen and/or 75 casein) origin have been used for these purposes (Barbabietola et al. 2016; Ferrari et al. 2017). Both 76 77 animal and vegetal glues are made up of natural polymers, that is mainly proteins (i.e. collagen derived from the bones, skins, tendons and cartilage of mammalians or fish swimming bladder) or 78 79 polysaccharides (i.e. amylose and amylopectin derived from different plants, such as potato, rice, corn or wheat), respectively (Barbabietola et al. 2016; Ferrari et al. 2017). Through aging, these glues 80 undergo stiffening and thickening which may in turn generate distortions, tensions and discoloration, 81 or form intricate layers that are very recalcitrant to being removed (Blüher et al. 1995; Gostling 1989). 82 In the case of animal glues, humidity, temperature, UV radiation and pollutants can generate protein 83 cross-linking and/or hydrolysis/oxidation of peptide bonds, while microbial metabolism produces 84 acid molecules and pigmented spots (Barbabietola et al. 2016). Starch glue has been commonly used 85 for ancient textile restoration (Ahmed and Kolisis 2011; Whaap 2007). After aging, starch paste is 86 generally found in shrunk, cracked, rigid and brittle form, which cannot provide enough adhesion for 87 88 effective support. In this form, it can cause heavy damage to ancient textiles because of concomitant embrittlement, hardening, yellowness and acidity of the latter. Furthermore, starch may be a source 89 of contamination by amylolytic fungi and bacteria that contribute to textile decay over time, especially 90 when a suitable degree of humidity supporting microbial growth is present. 91

Consequently, cleaning of glue residues is often a priority in the restoration of ancient paper 92 or textile artworks. Current mechanical and chemical methods display serious drawbacks mainly 93 related to aggressiveness towards material and/or toxicity for the restorers and/or the environment. 94 Humidification (also called wet-cleaning) has been used to swell starch paste, however it generally 95 needs long treatments which are unsuitable for paper or textile artifacts and is often insufficient for 96 aged or hardened glues. Bio-based methods, i.e. the use of enzymes or microorganisms, have been 97 shown to be a very efficient alternative in a number of cases (Ahmed and Kolisis 2011; Barbabietola 98 99 et al. 2016; Ferrari et al. 2017).

Enzymes are certainly the most frequently used method for the treatment of glue-damaged 100 paper (Banik et al. 2003; Corbi et al. 2005; DeSantis 1983; Sandrine 2002) or textiles (mainly linen, 101 silk and cotton fabrics, so far) (Ahmed and Kolisis 2011; Ciatti et al. 2010) and several successful 102 examples have been reported in the literature. For instance, trypsin has been used for detaching a 103 compact block of leaves (Wendelbo 1976). Amylases and proteases have been employed for 104 detaching graphics from their backings (De la Chapelle 2003; Segal and Cooper 1977). Very recently 105 106 enzyme extracts with protease activity isolated from marine invertebrates have been used to remove aged/altered protein glue layers from the velinatura (Japanese paper bonded by animal glue) of 107 108 ancient oil on canvas or from polychrome wood (Palla et al. 2016). This proved to be a cutting-edge strategy also useful to bio-clean fragile artworks such as wax sculptures. An additional advantage of 109 these marine invertebrate-derived extracts is their antimicrobial activity useful to control 110 bacteria/fungi growth (Palla and Barresi 2017). It is worth noting that application of enzymes in 111 solution is not always suitable for paper or textile artworks, since it may involve artifact flooding with 112 excess water which favors mold and fungi colonization and growth and thus causes further damage 113 to the artifact (Ahmed and Kolisis 2011). Actually, water-dissolved α -amylase preparations have been 114 applied locally either in solution (Ahmed and Kolisis 2011) (Fig. 1a-c; Table 1) or as poultice (Bott 115 1990; Chapman 1986; Shibayama and Eastop 1996) for the bio-restoration of starch-glue treated 116 textiles. In general, the use of immobilized enzymes is preferable for these applications. Theoretically, 117 all enzymes can be immobilized. However, it is worth reminding that the immobilization yield and 118 the enzyme efficiency should be determined for each specific enzyme and immobilization strategy, 119 to limit the loss of enzyme and catalytic activity. A ready-to-use poultice of amylolytic enzymes, 120 121 called Albertina Kompresse, was developed by an Austrian group for removing non-swellable starchbased glue from graphic artworks of albums of the "Albertina" graphic collection in Vienna (Schwarz 122 et al. 1999). PhytagelTM was used for lowering and controlling water content in enzyme solutions 123 (Iannucelli and Sotgiu 2009) used for cleaning -etchings depicting the Chinea of Clemente VIII, 124 dating 1598. Gellan hydrogel-immobilized α -amylases have been developed for removing starch 125 126 paste from ancient paper documents (Mazzuca et al. 2014). A gellan-immobilized bacterial α-amylase 127 has been recently used to clean a wool shroud dating back to the Coptic period from a starch glue that had been used in the 1950s to temporary consolidate the textile (Ferrari et al. 2017) (Fig. 1d-h; Table 128 1). After selection of the suitable enzyme (among those commercially available) and optimization of 129 the conditions for enzyme immobilization, the cleaning of the back of the two fragments (about 4 m² 130 of textile) composing the tunic was completed in 160 h of work (Ferrari et al. 2017). A recent study 131 (Barbabietola et al. 2016) has described the first attempt to bio-cleaning ancient paper from animal 132 133 glue by using living bacteria (Table 1). To this aim, non-pathogenic, non-spore-forming and noncellulolytic *Ochrobactrum sp.* TNS15E was used after immobilization on agar gel. This bacterial pack
 was used to remove glue layers from paper documents dating back to the 17-18th century. Four-hour
 treatment was sufficient to clean the cellulose fibers from glue, as confirmed by both colorimetric
 and scanning electron microscopy (SEM) analyses.

Apart from paper/textile biocleaning from glue, it is worth reminding the case of aged dryingoil stains on both ancient paper and textiles. During drying and aging, double bonds of unsaturated fatty acids are oxidized by oxygen in the air, giving rise to multiple products which include nonhomogeneous polymeric network of triacylglycerides which may be hard to being removed (Ahmed et al. 2010; Blüher et al. 1997). Lipases such as that of *Candida cylindracea* have been used to clean such aged drying-oily stains from paper documents (Blüher et al. 1997) or textiles such in the case of a coptic tunic (Ahmed et al. 2010) (Table 1).

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146 **Biorestoration/biocleaning of stone artworks**

Durable stone (e.g. marble and/or limestone) has been used for the construction of a multitude 147 of artworks and monuments all along the human history and all over the world including the Egyptian 148 Pyramids, the Greek and Roman temples and theaters, the European Cathedrals and the Taj Mahal in 149 India. Unfortunately, all of them have been suffering from progressive deterioration caused by both 150 biotic and abiotic agents (Dhami et al. 2014). These numerous factors have led to stone dissolution, 151 staining or color alteration, surface alteration, bio-corrosion and transformations into smaller sized 152 crystals, etc... (Chand and Cameotra 2011). In the recent decades, microbial biofilm production, 153 deposition of organic (such as residual hydrocarbons and other organic pollutants in dust) and 154 inorganic compounds (formation of nitrate and sulfate alterations such as the black crusts) have been 155 among the main deterioration events (Antonioli et al. 2005; Di Pippo et al. 2009; Fernandes 2006; 156 Warscheid and Braams 2000). Actually, limestone mainly consists of the most stable polymorph of 157 calcium carbonate, i.e. calcite, (with only a small content of aragonite) but is very porous and 158 hydrophilic. This makes limestone very susceptible to water flush (especially acid rain), 159 environmental pollutants and physical, chemical and biological (e.g. microorganisms) weathering 160 (Dhami et al. 2014). Therefore, survival of many cultural and historical assets is in threat. One of 161 such examples is the cave of Lascaux in southwest France which is considered the best conserved 162 prehistorical example of human wall painting art (they are also named the paleolytic Cappella 163 Sistina). In this site, infection of *Fusarium sp.* and other molds have deteriorated the floor and banks 164 of the main chamber (Rosenbaum 2006), but also autotrophic organisms such as green algae have 165 produced green pigments because of the intense illumination and improved CO₂ availability related 166

167 to visitors (Bastian et al. 2010). Martin-Sanchez and co-workers (2012) have extensively studied the effectiveness of biocides in the cave biocleaning. Fungicides had been intensively applied to treating 168 the cave since 2001 that were essentially targeted to remove *Fusarium sp* but obtained little success. 169 In 2008, a new biocide treatment was planned due to black stains that appeared on the cave surfaces. 170 DGGE analysis on these stains showed the presence of Ochroconis lascauxensis. This result 171 172 demonstrated the ineffectiveness of the previous biocide treatments on the long time which appeared 173 to favor colonization by other fungal strains and therefore increase fungal diversity. Later, i.e. in 2010, fungal communities were quite different from those detected in 2008, since the main identified 174 strain was a yeast belonging to the *Herpotrichiellaceae* family. It is clear that careful preliminary 175 study on the possible advantages and disadvantages of applying biocides in subterranean 176 environments is required (Martin-Sanchez et al. 2012). 177

Many attempts have been made to fix such structural damages by application of traditional 178 conservative treatments such as organic and inorganic chemicals (Lazzarini and Laurenzi Tabasso 179 1986). However, these agents have been most often low effective, in spite of their aggressiveness 180 181 (which, on the other side, has led the concomitant risk of further damaging the artwork). Moreover, 182 these strategies involve the use of high amounts of solvents, which are finally discarded in the environment creating problems of sustainability (Dhami et al. 2014). Alternatively, physical 183 184 treatments such as laser cleaning have been used, but at significantly higher costs (Germinario et al. 2017). Furthermore, all these treatments have short duration effects thus requiring repeated 185 186 interventions with relevant economic issues for public and private conservation agencies. Overall, conventional treatment methods have therefore proved to be unsatisfactory. 187

The shortcomings of conventional strategies have encouraged research in new conservation 188 and remediation strategies based on biological methods (Fernandes 2006). As for the treatment of 189 other type of artifacts, bio-based restoration approaches for stone materials are characterized by lower 190 cost, toxicity and aggressiveness towards the artworks (Germinario et al. 2017). As described in the 191 following sections, bio-based methods have been used to remove different degradation products from 192 stone monuments, wall paintings, and marble statues (Germinario et al. 2017), including deposits of 193 environmental pollutants (Margesin et al. 2011) and synthetic polymers present in adhesives 194 195 (Giordano et al. 2018) as well as in paints used by graffiti writers (Sanmartin et al. 2014) (Fig. 2; Table 1). In addition, biocleaning has been performed on stone artworks suffering from inaccurate or 196 aged restoration intervention (Beutel et al. 2002; Antonioli et al. 2005) (Fig. 2; Table 1). 197

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199 <u>Removal of sulfate and nitrate alterations</u>

One of the most important causes of decay of calcareous stones is the conversion of calcium 200 carbonate into calcium sulfate (gypsum) mainly caused by acid rains (i.e. containing significant 201 amounts of sulfuric and nitric acid) (Ranalli et al. 1997). For instance, the genesis of "gypsum crusts" 202 on the surface of such porous material can engender following fractures of the underlying stone. 203 204 When calcium sulfate salts are accumulated together with atmospheric particles (pollen, dust, spores, small particles of smog) the so called "black crusts" are formed (Fig. 2a). For the removal of sulfates 205 from artistic stoneworks, procedures based on the use of sulfate-reducing bacteria have been reported. 206 207 Different bacterial strains of the genus Desulfovibrio (e.g. D. desulfuricans and D. vulgaris) (either pure or in mixed cultures) have been applied under anaerobic conditions to marble samples directly 208 or after adhesion to a sepiolite matrix (Ranalli et al. 1997). The use of sepiolite promoted sulfate 209 removal on both simulated samples and real marble statue artifacts. On the latter, 81 % sulfate 210 removal was obtained after 36 h treatment (Ranalli et al. 1997). Actually, D. desulfuricans (Ranalli 211 212 et al. 1997) and D. vulgaris (Ranalli et al. 1997; Cappitelli et al. 2007a; Alfano et al. 2011) have been widely employed in restoration/removal of sulfate crusts from other artifacts (Table 1). Use of 213 biotechnological cleaning on durable stone monuments can sometimes comply with multiple types 214 of deterioration such as described by Cappitelli et al. (2007b). In this study, the sulfate-reducing 215 bacterium D. vulgaris subsp. vulgaris ATCC 29579 was employed to remove the black crust found 216 on marble of the Milan Cathedral (Italy). Compared to chemical cleaning (i.e. ammonium carbonate-217 EDTA) strategy, the microbial-catalyzed approach resulted in more homogeneous removal of the 218 219 deposits and higher preservation of the original surface (Cappitelli et al. 2007b) (Table 1). Both 220 chemical and biological treatments converted gypsum (i.e. calcium sulfate) to calcite (i.e. calcium 221 carbonate), allowing consolidation. However, the chemical strategy also formed undesirable sodium sulfate while the use of *D. vulgaris* did not (Cappitelli et al. 2007b). Nonetheless, biological removal 222 223 of sulfates may require quite long application periods, depending on the thickness of the crust. A recent study has demonstrated that this period can be greatly shortened and general efficiency of 224 biocleaning can be significantly improved by combining the use of sulfate-reducing bacteria with a 225 226 non-biological strategy, e.g. the use of a non-ionic detergent (Troiano et al. 2013) (Fig. 2a, b; Table 227 1). This combined strategy shortened application times of about 38-70 % depending on the specific artifact to be cleaned (Troiano et al. 2013). 228

Another consequence of acid rains (and of the action of living microorganisms) is the deposit of calcium nitrate salts on stone buildings and wall paintings (Dhami et al. 2014). Here, again, pollution increases the presence of various nitrogen oxides in the atmosphere that in turn may react with rain water and form nitrous and, more abundantly, nitric acid which then reacts with stone and replaces calcium carbonate with calcium nitrate (Dhami et al. 2014). Different strains of

Pseudomonas spp. have been recently applied for removing calcium nitrate salts from two stone 234 monuments. Agar-entrapped Pseudomonas stutzeri DSMZ 5190 has been used for the biocleaning of 235 nitrate efflorescence from wall paintings located in the lunettes of the central vault of the Santos 236 Juanes church in Valencia, Spain (Bosch-Roig et al. 2013) (Fig. 2c-e; Table 1). The chosen strategy 237 proved to be extremely efficient allowing to remove 92 % of the precipitates in 90 minutes. 238 Pseudomonas pseudocaligenes KF707 has been used to remove nitrate salts from the tuff stone 239 surfaces of the 12th century Matera Cathedral, Italy (Alfano et al. 2011) (Table 1). Here, carbogel-240 entrapped bacteria were applied to the Cathedral walls and allowed quick removal of the surface 241 242 nitrate deposits, since 55 % of the nitrate salts were "cleaned" after 24 h.

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244 <u>Bioconsolidation</u>

245 Apart from interventions aimed at removing superficial deposits and/or crust from stone monuments, the use of calcifying bacteria offers a chance to consolidate decayed building structures 246 247 and materials. This application, sometimes also called microbial geotechnology (intending microbialbased technology for civil structures) actually mimics nature since many carbonate rocks have been 248 249 cemented by carbonate precipitation induced by microorganisms during Earth geological cycles. This relatively novel and environmental-friendly technology has been studied for at least 20 years and has 250 already been used for protecting and/or restoring different decayed construction materials/artifacts 251 252 (Dhami et al. 2012; 2013).

Calcium carbonate precipitation is a chemical process (described by equation 1) which is influenced by four main factors, i.e. calcium concentration, amount of dissolved inorganic carbon (DIC), availability of nucleation sites and pH (Hammes and Verstraete 2002).

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$$Eq.1 \qquad \qquad Ca^{2+} + CO_3^{2-} \leftrightarrow CaCO_3$$

257
$$Eq. 2$$
 $K_{SP,Calcite,25^{\circ}C} = [Ca^{2+}][CO_3^{2-}] = 4.8 \times 10^{-9}$

Calcium carbonate precipitation occurs when the product of concentrations of Ca^{2+} and CO_3^{2-} is 258 higher than the solubility product (K_{SP}) of calcium carbonate (Eq. 2). However, the amounts of CO_3^{2-1} 259 260 in a given system depends on both the amount of DIC (which in turn depends on several parameters such as temperature and partial pressure of carbon dioxide) and pH. Because of the high number of 261 262 parameters that may contribute to control calcium carbonate precipitation, different bacteria, isolated from different habitats, are able to create local micro-environments that induce such phenomenon 263 (Hamilton 2003) (Fig. 3). The four main groups of microorganisms that may influence calcification 264 are: (i) photosynthetic organisms such as cyanobacteria and algae, (ii) sulfate reducing bacteria 265

responsible for dissimilatory reduction of sulfates, (iii) organisms utilizing organic acids, and (iv) 266 organisms that are involved in nitrogen cycle either by ammonification of amino acids/nitrate 267 reduction or hydrolysis of urea (Stocks-Fischer et al. 1999; Hammes and Verstraete 2002; Jargeat et 268 al. 2003) (For an exhaustive review please refer to the study of Dhami et al. 2014). The precipitation 269 of carbonates by bacteria through urea hydrolysis is the most straightforward and easily controlled 270 mechanism of microbial induced calcium carbonate precipitation since it produces high amounts of 271 carbonates and an alkaline environment (Dhami et al. 2014). Boquet et al. (1973) firstly demonstrated 272 273 the precipitation of calcium carbonate by soil bacteria under laboratory conditions (Fig. 3). At that 274 time, several Bacillus spp. and Pseudomonas aeruginosa were shown to form calcite crystals. In 1990, Adolphe et al. patented the concept of using calcifying microorganisms to treat artificial 275 276 surfaces and founded the "Calcite Bioconcept" company. However, the first in situ application of bioconsolidation was carried out in Thouars (France) on the tower of the Saint Médard Church by 277 278 using Bacillus cereus only in 1993 (Le Metayer-Levrel et al. 1999) (Table 1). Although this application was judged as successful, some drawbacks were: the need to regularly repeat the treatment 279 280 (for instance, each 10 years); the presence of natural pigments in the nutritional medium of *B. cereus* which co-precipitated with calcium carbonate thus giving the new stone layer a light persistent 281 coloring; the formation of endospores and a thin biofilm of Bacillus sp. For these reasons, Rodriguez-282 Navarro et al. (2003) proposed to replace Bacillus sp. with a Gram-negative, non-pathogenic soil 283 bacterium, i.e. Myxococcus xanthus. Tiano et al. (1999) studied the effect of Micrococcus spp. and 284 Bacillus subtilis on Pietra di Lecce bioclastic limestone. Variations in the kind of bacteria used and 285 the methods for bacterial cell delivery to the stone surface have been tested by different Authors with 286 variable success (Daskalakis et al. 2014; Dhami et al. 2014; Helmi et al. 2016; Micallef et al. 2016). 287 Recently, the use of indigenous calcifying bacteria for re-inoculation of stone monuments has been 288 proposed as an alternative strategy for bioconsolidation (Jroundi et al. 2017). However, 289 microbiologically driven calcification remains more complex than chemical methods, since microbial 290 activity depends on many factors such as temperature, pH, concentrations of donors and acceptors of 291 292 electrons and concentration and diffusion rates of nutrients and catabolites. Hence, the use of 293 microbial calcification at large scales has not been always encouraged since it may be hard to manage (Dhami et at. 2014). Also the cost of media required for bacterial growth may be a significant 294 295 economic limit of this approach (Achal et al. 2009; 2010).

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297 <u>Biorestoration of wall paintings</u>

As previously mentioned, also stone artworks may suffer from inaccurate or aged restoration 298 strategies. This was the case of two wall paintings covered by animal glue layers during restoration 299 interventions which have been restored using different bio-based approaches as described by Beutel 300 et al. (2002) and Antonioli et al. (2005), respectively (Table 1; Fig. 2). The first study concerns 301 medieval wall paintings called "Falcon hunt-meeting of the living and the dead" located in St. 302 Alexander church in Wildeshausen, Germany. These paintings were suffering from severe peeling 303 off from the roughcast surface (Beutel et al. 2002) (Table 1). Actually, this is a common problem of 304 305 wall paintings in medieval churches of the Northern Europe since they have long been treated by 306 application of casein layers to stabilize them (Beutel et al. 2002). As for other glue-like matrices, aging in addition to climate effects causes progressive hardening and stiffening of casein layers thus 307 causing an even more drastic peeling off of the painted parts from the surface (Beutel et al. 2002). 308 This problem was fixed by removing aged casein layers through application of a selected microbial 309 310 serine-protease (Alcalase 2.5 DX-L). The enzyme was covalently immobilized onto an epoxidefunctionalized cellulose acetate membrane (Beutel et al. 2002). By 2D fluorescence monitoring of 311 the tryptophan exposed by casein hydrolysis, it could be estimated that 30-minute treatment was 312 sufficient for substantial removal of the casein layers from the mural painting. 313

The case of "Conversion of S. Efisio and battle" fresco by Spinello Aretino at the monumental 314 315 Cemetery of Pisa (Italy) was even more complicated (Fig. 2f, g; Table 1). Because of weathering and other environmental aging, this fresco needed to be restored and for this purpose it was removed from 316 317 the wall surface by using the tear-off technique in the 1980s. Firstly, the fresco was covered with a gauze that was sticked by applying an animal glue (mixed with high concentrations of formaldehyde, 318 319 as antimicrobial agent). Once the glue had been hardened, the fresco was detached from the wall. Unfortunately, the fresco was then forgotten until 2000s in a storeroom, so that, traditional application 320 321 of protease mixtures were unable to remove the gauze (Antonioli et al. 2005). Because of the long time of storage, it is likely that the presence of formaldehyde had promoted the formation of a resistant 322 net of cross-linked proteinaceous material, which was very recalcitrant to protease catalysis. 323 However, the selection of a bacterial strain (i.e. Pseudomonas stutzeri) able to grow on chips of 324 insoluble glue harvested from the "clothed" fresco (and hence possessing the right collagenase 325 enzymes), finally helped to solve the problem. Actually, it was then possible to use this bacterial 326 strain (i.e. cotton strips impregnated with live Pseudomonas stutzeri were applied) directly on the 327 fresco and completely degrade the glue layer and remove the cloth from the fresco after only 12 hour 328 treatment (Fig. 2f, g). 329

331 <u>Biocleaning of graffiti and synthetic adhesives</u>

Stone monuments are not only aggressed by atmospheric events or microorganisms but, unfortunately, also by vandalism acts, including painting by unauthorized graffiti. Graffiti materials have a complex chemical composition that comprises synthetic polymers such as acrylics, alkyds and nitrocellulose, and several additives (Germinario et al. 2017). Quick removal of graffiti is an important issue, since the fresher the graffiti, the easier is their removal. Here again, bio-based (through enzymes or microorganisms) removal of synthetic materials is an emerging strategy which has already shown good results in a number of cases.

One of the first examples of acrylic material removal by means of bio-based approach has 339 been described by Bellucci et al. (1999). Here, a lipase has been used to eliminate aged acrylic 340 ParaloidB72 resin from a 15th century tempera painting on panel and a 19th century oil painting on 341 canvas. In both artworks, the presence of surface layers containing ParaloidB72 was the result of 342 previous restoration interventions occurred in the early 1970s and in the 1980s, respectively (Bellucci 343 344 et al. 1999). Cleaning was likely achieved via hydrolysis of the ester groups of the acrylate and methacrylate units contained in the synthetic resin leading to free carboxylic acid groups. This 345 346 reaction therefore generated more hydrophilic products which facilitated acrylic resin removal by aqueous cleaning systems (Bellucci et al. 1999). Bio-based removal of acrylic materials is particularly 347 348 advantageous for treating painting where the use of traditional methods, e.g. organic solvents, would likely remove also original paint layers. However, the same approach can be employed also to treat 349 350 stone materials. For instance, Germinario et al. (2017) tested different lipases in oil-in-water micro-351 emulsions for the removal of acrylic marker pen inks from unglazed ceramic surfaces. Very recently, Palla and co-workers (2017) successfully detached a canvas layer glued by the same acrylic 352 ParaloidB72 resin to a mosaic sample by applying a gelled (3% Klucel G) enzymatic solution with 353 esterase activity. The bio-removal was very fast, only 40 minutes at room temperature (Palla et al. 354 2017). The same research group also reported the removal of adhesive-tape glue residues present on 355 specific areas of an acrylic paint on canvas. They used a microemulsion of Velvesil Plus® (a 356 surfactant used in the cosmetic field) containing an esterase derived by a marine organism that proved 357 to be active even at a temperature lower than 30°C (whereas most commercially available enzymes 358 359 have an optimum temperature of 37°C) (Palla et al. 2016). These authors demonstrated that the enzymatic solution can be merged into the Velvesil Plus® gel (without any negative effect on enzyme 360 activity) and easily applied to remove the undesired layers. The contact between enzyme and the layer 361 362 to be removed was obtained by gently moving the microemulsion, by a soft-brush for 5 minutes 363 (Giordano et al. 2018). As regards the use of living microorganisms, D. desulfuricans has proved able

to degrade nitrocellulose-based paints (Giacomucci et al. 2012), while the use of different bacterial
strains has been tested for the bio-cleaning of acrylic polymers used in the restoration field (Troiano
et al. 2014).

367 **CONCLUSION**

This report demonstrates that it is possible to face cultural heritage damage due to aging, 368 weathering, pollution or wrong restoration interventions by using bacteria or purified enzymes 369 370 suitably immobilized to contain the risk of employing aqueous solutions. Among the wide range of enzymes commercially available those displaying a good catalytic activity at low temperatures (lower 371 than 30° C) are very promising since they can be applied also to fragile items. The microbial world as 372 373 well as the marine environment seem to be good candidates to be explored for finding such enzymes. This field is promising also to find, in the future, a solution to contain microbial deterioration (Mazzoli 374 et al. 2018) thus avoiding the use of acids, solvents and surfactants (dangerous for the artworks, the 375 376 art restorers and the environment) for instance by using enzyme- or bacteriocin-mediated bacterial competition. In this case, the safety and effectiveness of the microorganisms employed is mandatory 377 and the need for control and analysis before and after treatments strongly recommended. 378 Interdisciplinary approaches and collaborations between art conservators and biotechnologists, 379 biochemists and microbiologists is the essential requisite to preserve objects that state the immense 380 381 creativity of artists and the high value of human history.

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383 COMPLIANCE WITH ETHICAL STANDARDS

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Figure Legends

Fig. 1. Biocleaning of ancient textiles from starch glue. Pictures refer to biocleaning of a historical carpet dating back to the Ottoman period and exhibited in the museum of the Faculty of Applied Arts, Helwan University, Egypt (a, b, c) (modified from Ahmed and Kolisis 2011) and a coptic tunic dating back to the 5-6th century A.D. and exposed at the Egyptian Museum, Turin, Italy (d, e, f, g, h) (modified from Ferrari et al. 2017). b, c Detail of the carpet before (b) and after (c) the α -amylase treatment. e-h, Details of the Coptic tunic before (e, g) and after (f, h) the α -amylase treatment.

Fig. 2. Biorestoration of stone monuments. a, b, Detail of a marble statue dedicate in 1921 by Lina Arpesani to the poetess Anna Zuccari and located in the Monumental Cemetery of Milan (Italy). The black crusts (a) affecting the statue were cleaned (b) by using sulfate-reducing *Desulfovibrio vulgaris* (modified from Troiano et al. 2013). c, d, e Cleaning of the wall painting in the lunette of the Santos Juanes church, Valencia, Spain from nitrate salt efflorescence by means of agar gel-entrapped *Pseudomonas stutzeri*. Pictures represent the fresco area before (c), during (d) and after biocleaning (modified from Bosch-Roig et al. 2013). f, e biorestoration of the Spinello Aretino fresco "Conversion of S. Efisio and battle" in the Monumental Cementery of Pisa (Italy). f For animal glue removal. Cotton strips impregnated with live *Pseudomonas stutzeri* were applied leading to fresco biocleaning (g) (modified from Antonioli et al. 2005)

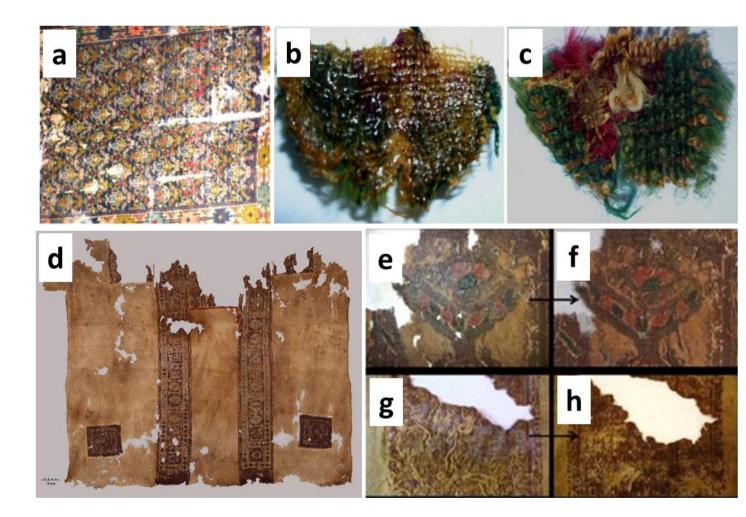
Fig. 3. Calcifying bacteria. Colonies of 6 different strains of *Bacillus sphaericus* and *Bacillus lentus* on agar plates during calcium carbonate deposition are shown (Dick et al. 2006).

Type of	Specific artwork (specimen)	Historical period of	Issue	Biorestoration/biocleaning	Reference
artwork		the specimen		strategy	
Paper	Graphic artworks from albums (Graphic	XIX century A.D.	Removal of aged	Gel-entrapped α-amylase	Schwarz et
	Collection Albertina, Vienna, Austria)		starch glue		al. 1999
Paper	Paper documents of the Genoese Republic	XVII-XVIII	Removal of aged	Agar-immobilized	Barbabietola
	(Central Institute for Graphic Arts, Rome,	century A.D.	animal glue	Ochrobactrum sp. TNS15E	et al. 2016
	Italy)				
Textile	Coptic tunic (Greek-Roman Museum,		Removal of aged	Lipase from <i>Candida</i>	Ahmed et al.
	Alexandria, Egypt)		oily stains	cylindracea	2010
Textile	Carpet (Museum of the Faculty of Applied	Ottoman period	Removal of aged	α-amylase from Aspergillus	Ahmed and
	Arts, Helwan University, Egypt)	_	starch glue	oryzae	Kolisis 2011
Textile	Coptic tunic (Egyptian Museum, Turin,	V-VI century A.D.	Removal of aged	Gellan immobilized α-	Ferrari et al.
	Italy)		starch glue	amylase from <i>Bacillus sp</i> .	2017
Stone	Milan Cathedral (Italy)	XV century A.D.	Removal of black	Sulfate-reducing	Cappitelli et
monument			crust	Desulfovibrio vulgaris	al. 2007b
				ATCC 29579	
Stone	Florence Cathedral (Italy)	XV century A.D.	Removal of black	Carbogel-entrapped sulfate-	Gioventù et
monument	-		crust	reducing <i>Desulfovibrio</i>	al. 2011
				vulgaris ATCC 29579	
Stone	Matera Cathedral (Italy)	XII century A.D.	Removal of	Carbogel entrapped nitrate-	Alfano et al.
monument		•	nitrate and	reducing <i>Pseudomonas</i>	2011
			sulphate crusts	pseudoalcaligenes KF707	
			1	and sulfate-reducing	
				Desulfovibrio vulgaris	
				ATCC 29579	
Stone	Stone column and marble statue,	XX century A.D.	Removal of black	Desulfovibrio vulgaris	Troiano et al.
monument	Monumental Cemetery (Milan, Italy)	•	crust	ATCC 29579 plus non-ionic	2013
				detergent	

Table 1. Some of the most significant examples of biorestoration/biocleaning of artworks described in the present study.

Stone	Saint Médard Church, (Thouard, France)	XII century A.D.	Limestone	Bacillus cereus	Le Metayer
monument			bioconsolidation		et al. 1999
Wall	Wall paintings of the lunettes of the central	XVII-XVIII	Removal of	Agar-entrapped	Bosch-Roig
paintings	vault, Santos Juanes church (Valencia,	century A.D.	calcium nitrate	Pseudomonas stutzeri	et al. 2013
	Spain)		salt efflorescence	DSMZ 5190	
Wall	Falcon hunt-Meeting of the living and the	XIV century A.D.	Removal of aged	Covalently immobilized	Beutel et al.
paintings	dead, St. Alexander church (Wildeshauen,		casein layers	protease (Alcalase 2.5 DX-	2002
	Germany)			L)	
Wall	Conversion of S. Efisio and battle by	XIV century A.D.	Removal of aged	Cotton strips impregnated	Antonioli et
paintings	Spinello Aretino, Monumental Cemetery		formaldehyde-	with Pseudomonas stutzeri	al. 2005
	(Pisa, Italy)		reated animal glue	A29	
Painting	The Visitation with St. Joseph, St. Zacharias	XV century A.D.	Removal of	Lipase from <i>Candida</i>	Bellucci et
on panel	and Four Angels		acrylic resin	cylindracea	al. 1999
Painting	Portrait of a man	XIX century A.D.	Removal of	Lipase from <i>Candida</i>	Bellucci et
on canvas			acrylic resin	cylindracea	al. 1999







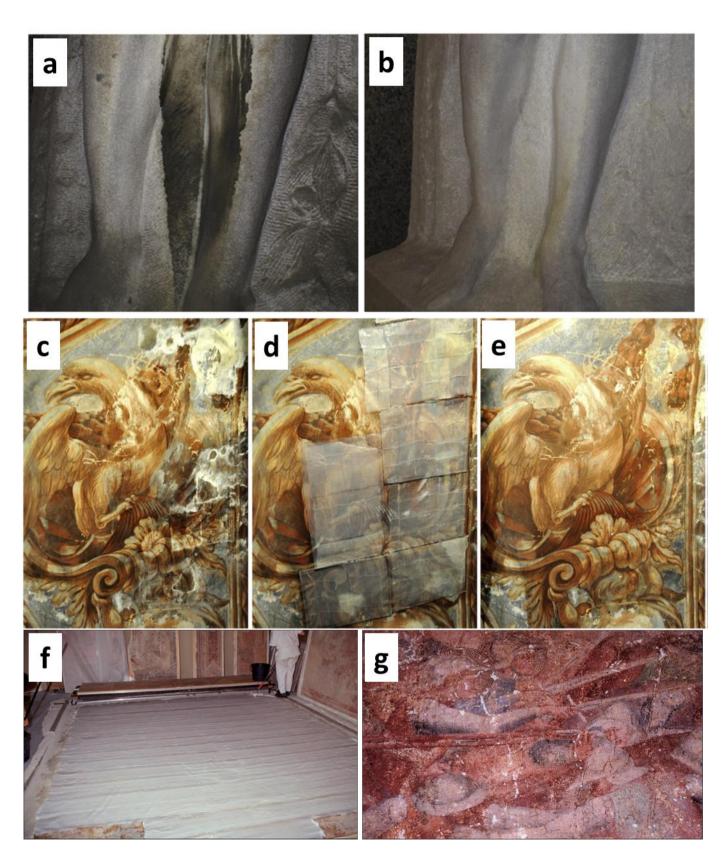


Fig. 3.

